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The pp24 phosphoprotein of Mason-Pfizer monkey virus contributes to viral genome packaging

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Abstract

Background: The Gag protein of Mason-Pfizer monkey virus, a betaretrovirus, contains a phosphoprotein that is cleaved into the Np24 protein and the phosphoprotein pp16/18 during virus maturation. Previous studies by Yasuda and Hunter (*J. Virology*. 1998. 72:4095–4103) have demonstrated that pp16/18 contains a viral late domain required for budding and that the Np24 protein plays a role during the virus life cycle since deletion of this N-terminal domain blocked virus replication. The function of the Np24 domain, however, is not known.

Results: Here we identify a region of basic residues (KKPKR) within the Np24 domain that is highly conserved among the phosphoproteins of various betaretroviruses. We show that this KKPKR motif is required for virus replication yet dispensable for procapsid assembly, membrane targeting, budding and release, particle maturation, or viral glycoprotein packaging. Additional experiments indicated that deletion of this motif reduced viral RNA packaging 6–8 fold and affected the transient association of Gag with nuclear pores.

Conclusion: These results demonstrate that the Np24 domain plays an important role in RNA packaging and is in agreement with evidence that suggests that correct intracellular targeting of Gag to the nuclear compartment is an fundamental step in the retroviral life cycle.

Introduction

Viruses of the *Betaretroviruses* genus, formerly known as D- and B-type retroviruses, assemble their capsids in the cytoplasm of infected cells instead of at the plasma membrane like most retroviruses. The B-type viruses contain prominent surface glycoproteins and spherical, eccentric capsids and include mouse mammary tumor virus (MMTV) and exogenous and endogenous MMTV-like retroviruses in mice and humans [1-3]. D-type viruses have less dense surface spikes and contain cylindrical capsids. Exogenous and endogenous D-type viruses infect in a variety of mammalian hosts including Old World monkeys (Mason-Pfizer monkey virus [M-PMV], simian retrovirus 1 [SRV-

1], [SRV-2] and simian endogenous retrovirus) [4-6], New World monkeys (squirrel monkey retrovirus [SMRV]) [7], sheep and goats (Jaagsiekte sheep retrovirus and enzootic nasal tumor virus respectively) [8-10]. D-type virus sequences have also been detected in humans, the Australian common brushtail possum and mice (*Trichosurus vulpecula* endogenous retrovirus D, rabbit endogenous virus H, and MusD, respectively) [11-13].

M-PMV, the prototypical D-type virus, was first isolated from a mammary adenocarcinoma of a female Rhesus monkey [14]. Although M-PMV was originally suspected to be an oncogenic virus, it was later found to induce a

sever "wasting" and immunodeficiency syndrome distinct from that caused by immunosuppressive lentiviruses [15]. SRV-1 and SRV-2 are related to, yet serotypically distinct from, M-PMV and were isolated from primates suffering diseases similar to that caused by M-PMV [16,17].

M-PMV, the most thoroughly understood of the D-type betaretroviruses, contains four genes (5'-*gag-pro-pol-env*). As with other retroviruses, its Gag protein, Pr78, serves multiple functions during the viral life cycle, including virus assembly, virion maturation and early post-entry steps in virus replication [18]. Multiple studies have shown that Pr78 has the innate ability to assemble into immature capsids or procapsids in the cytoplasm, recognize and package the viral RNAs and glycoproteins and facilitate budding from the plasma membrane. During viral budding or shortly thereafter, Pr78 is cleaved by the viral protease to yield the mature virion associated proteins: matrix MA (p10), the phosphoprotein pp24, p12, capsid (CA or p27), nucleocapsid (NC or p14) and p4. These mature Gag-cleavage products then play roles during the early stages of the viral life cycle where they may help facilitate uncoating, reverse transcription and nuclear entry of the viral DNA. The regions and modifications of Pr78 required for these events have been partially identified.

Upon translation, Pr78 is targeted to a pericentriolar region of the cytoplasm in close proximity to the nuclear membrane where it assembles into spherical, procapsids [19]. The signal within Pr78 responsible for this pericentriolar targeting (the cytoplasmic targeting/retention signal or CTRS) is located within an 18 amino acid sequence of the matrix domain (MA). This motif is dominant over the bipartite myristylation and lysine/arginine-rich bipartite membrane targeting signals that is also located within the MA domain. Insertion of the CTRS into the analogous region of the MLV Gag protein, which normally assembles at the plasma membrane, results in intracytoplasmic assembly of MLV Gag. Secondly, substitution of an arginine within the CTRS of M-PMV Gag to a tryptophan (R55W) destroys the dominant CTRS function resulting in capsid assembly at the plasma membrane [20].

Other regions of Pr78 are also essential for procapsid assembly. Residues within the MA, yet separate from the CTRS, and the CA domains are required for assembly [20-23]. Likewise, the p12 domain within Pr78 provides an internal scaffolding that together with the conical I domain, which is located near the CA-NC junction, function to promote Gag-Gag interactions during capsid [24,25]. Assembly of the spherical capsid also requires interactions between the viral RNAs (vRNA) and Gag proteins [26,27]. Thus, the vRNA must also be present at the assembly site to provide this additional nucleation or scaf-

olding function. Although Gag-vRNA interaction occurs primarily through interaction in the NC domain, other regions of Gag appear to be important by targeting Gag to the site of vRNA packaging and by imparting correct structural information upon Gag [22,28-32].

Following assembly, the procapsids are transported to the plasma membrane from which they bud. Both the myristic acid modification of Pr78 and specific amino acids within the MA domain play critical roles in plasma membrane targeting [20,21]. Moreover, Sfakianos and Hunter have shown that the M-PMV Env glycoproteins and Rab11-positive recycling endosomes play critical roles in transporting the preassembled procapsids from the pericentriolar assembly site to the plasma membrane [33]. Upon arrival of the procapsids at the plasma membrane, a proline-rich PPPYX₄PSAP motif located near the carboxy-terminus of the Gag phosphoprotein, pp24, provides the late budding domain (L), which facilitates viral budding [34,35].

Upon release, nascent immature particles undergo a maturation process to acquire infectivity. During this process, Pr78 is cleaved by the viral protease to yield the seven proteins: MA, pp24, p12, CA, NC, and p4. The phosphoprotein conserved among M-PMV, SRV-1, SRV-2, SERV, and MMTV yet its function is only partially known. During M-PMV maturation, pp24 is further cleaved into two proteins; the C-terminal pp16 protein and the N-terminal Np24 protein. Both are required for virus replication. Yasuda and Hunter demonstrated that the pp16 domain contains the late budding motif and deletion of the Np24 domain completely blocked virus replication [35]. However, the function of Np24 was not determined. In this study, we examined the role of the Np24 domain during virus replication. We have identified a lysine-arginine rich motif within Np24 that is conserved among many betaretrovirus and is essential for infectivity. The results presented here show that the KKPGR motif in Np24 is not required for procapsid assembly, intracellular transport, budding or glycoprotein incorporation but plays a critical role in vRNA packaging.

Results

Deletion of the KR box in Np24 blocks virus replication

While it was previously determined that the Np24 domain of Pr78^{Gag} is required for replication [35], the role of this protein during the virus life cycle is not known. To gain further insight into which regions(s) of Np24 might be important for replication, the Np24 protein sequence was aligned to the analogous phosphoproteins from different infectious betaretroviruses (Fig. 1A). As expected because of the close similarity between the simian betaretroviruses M-PMV and SRV-1 and the more distantly related SRV-2 and SERV, their phosphoprotein

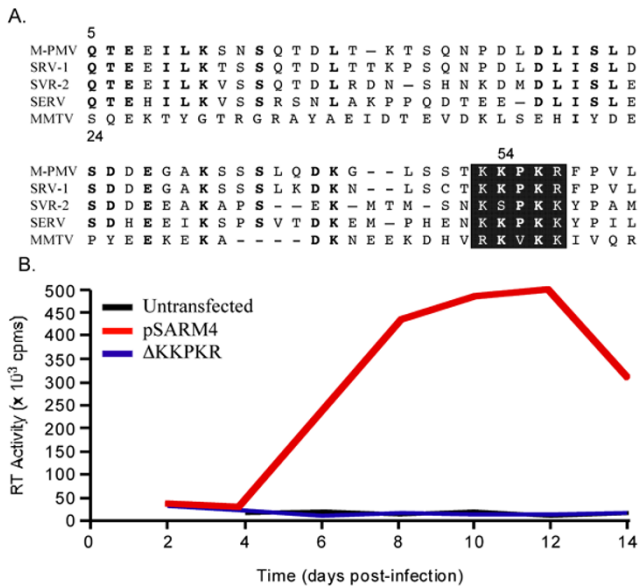


Figure 1
The highly conserved KR box within the Np24 protein of M-PMV is required for viral replication. (A) Amino acid alignment of phosphoproteins of betaretroviruses showing areas of sequence conservation in the N-terminus and C-terminus. All conserved residues are bolded and the highly conserved KR box is shadowed. Accession Numbers; M-PMV (P07567), SRV-1 (AAA47730), SRV-2 (P51516), MMTV (AAF3147) (B) Viral replication measured by virus spread assay. Culture media from COS-1 cells transfected with nothing (Black), wild-type M-PMV proviral DNA, pSARM4 (Red), or ΔKKPKR proviral DNA (Blue) were collected and assayed for RT activity. HOS cells were infected with equal amounts of virus (normalized by RT activity). Virus spread in HOS cells was measured by RT assays at 2, 4, 6, 8, 10, 12, and 14 days post infection.

sequences are 82%, 62%, and 61% (respectively) similar to M-PMV Np24. The most notable similarities occur at the amino- and carboxy-terminal ends of these phosphoproteins. While the amino-terminal sequences are not conserved in the phosphoproteins of MMTV and MMTV related betaretroviruses, the highly conserved cluster of positively charged amino acids, KKKPKR, (the KR box) near the carboxy-terminal end is shared by these betaretroviruses. In M-PMV (and SRV-1), the KR box is located near the carboxy-terminal end of Np24. In MMTV, it is located in the same relative position of pp21. The conservation of the KR box within the phosphoproteins of these divergent viruses suggests that it may be essential to virus replication. To determine if this motif (KKPKR) in M-PMV serves an important role during the virus life cycle, PCR mutagenesis was used to delete the region encoding the KKKPKR motif from the infectious clone pSARM4. The mutant, pΔKKPKR, and wild-type pSARM4 proviral DNAs were transfected into COS-1 cells. At 48 h post transfections the

viruses produced from the transfected cells were harvested and assayed for RT activity. Wild-type M-PMV and ΔKKPKR-transfected cells produced equivalent amounts of virus particles (data not shown). Viral spread assays were then carried out to examine if the deletion of the KKKPKR motif affected viral replication. For this, Hos cells were infected with equivalent amounts of wild-type and mutant virus particles, normalized by RT activities. The amounts of virus particles present in the supernatants of infected cells at 2, 4, 8, 10, and 12 days post-infection were determined by RT assays. While wild-type virus replicated in Hos cells, as indicated by the increasing amounts of RT activity in the culture medium over time, no detectable RT activity was observed in the supernatants of uninfected or ΔKKPKR-infected COS-1 cells even at 14 days post-infection (Fig. 1B). These data demonstrates that deletion of the KKKPKR motif blocked virus replication.

Assembly and release of ΔKKPKR mutant particles

Because the different morphogenic steps (procapsid assembly, cytoplasmic transport, membrane binding, and budding) are temporally separate for M-PMV, this virus provides an ideal opportunity to determine which if any of the late assembly steps might be affected by the deletion mutation. To determine if the replication-defective ΔKKPKR mutant could assemble intracellular procapsids, transfected cells were lysed in a TX-100 lysis buffer that does not disrupt assembled procapsids. The assembled procapsids were separated from the soluble unassembled Gag proteins in the cellular lysates by centrifugation through a 20% sucrose cushion. The pelleted proteins were solubilized directly in protein loading buffer, separated by SDS-PAGE, and immunoblotted using anti-Pr78 antibodies. As expected, wild-type Gag (Pr78^{WT}) was detected in both the soluble fraction (unassembled) and pelleted fractions (assembled procapsids) of the cellular lysates (Fig 2A. Lanes 1 and 2). The presence of the ΔKKPKR mutant Gag protein (Pr78^{ΔKR}) in the soluble and pelletable fractions (Fig. 2A lanes 4 and 5) indicates that the mutant Gag proteins assembled into procapsids.

The rate of assembly and release of capsids from Pr78^{WT} and Pr78^{ΔKR} expressing cells were analyzed by pulse-chase analyses to determine if the deletion had caused defects in virus assembly and release. Transfected COS-1 cells were pulse labeled with [³⁵S] methionine-cysteine for 30 min. and chased for 1, 2, 4, and 8 hours in complete growth medium. Cell associated and released-virus-associated proteins at each time point were analyzed by immunoprecipitation using rabbit anti-Pr78 antiserum (Fig. 2)

Similar levels of Gag (Pr78), Gag-Pro (Pr95), and Gag-Pro-Pol (Pr180) fusion proteins were synthesized during the pulse labeling by cells expressing wild-type M-PMV

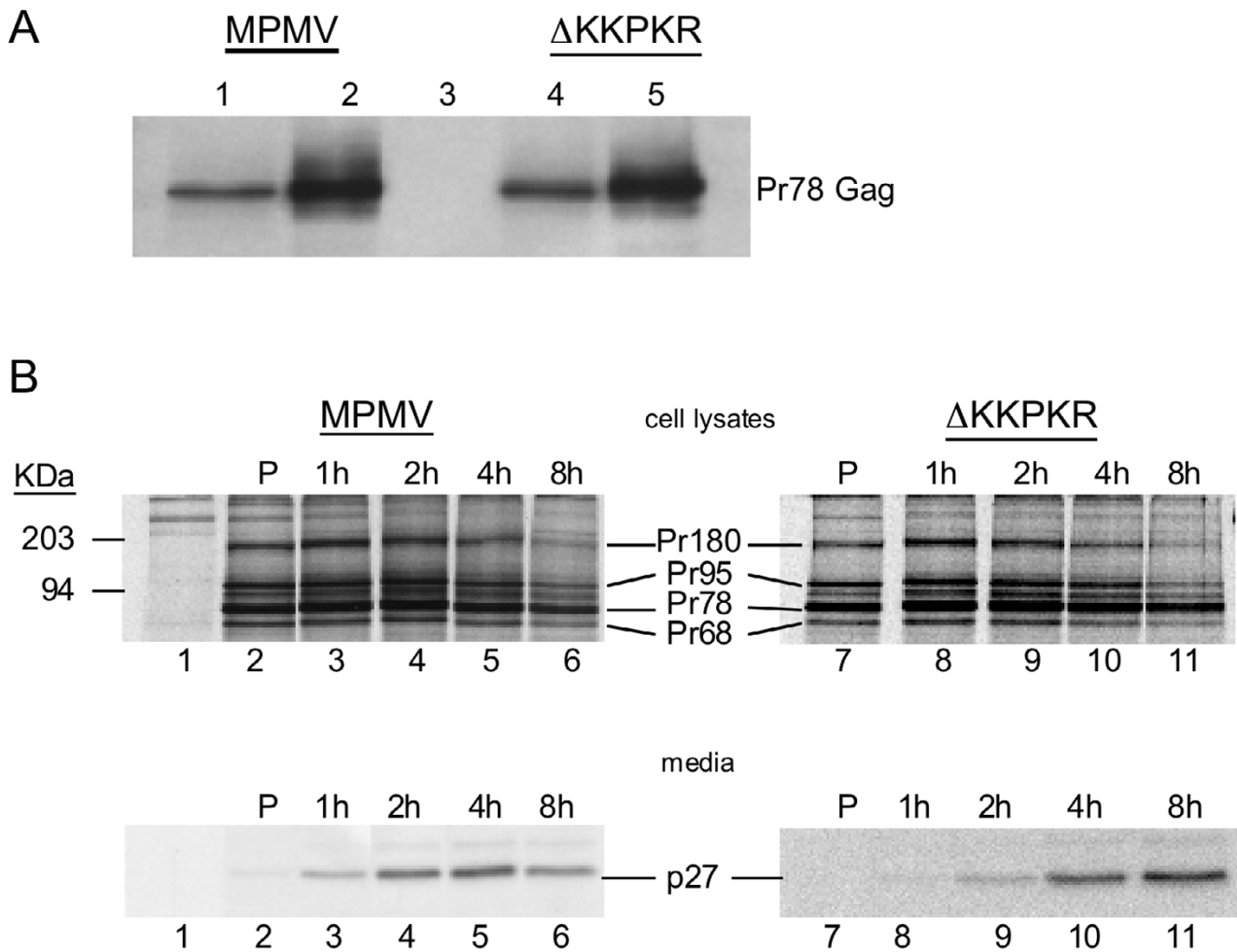


Figure 2
 (A) Western Blot analysis of intracellular procapsid assembly using Gag fractionation techniques. 48 hrs post transfection, COS-1 cells were lysed with and fractionated over a 20% sucrose cushion to separate assembled procapsids from unassembled Gag proteins. Pr78 in the fractionated samples were detected by western blot using rabbit anti-Pr78 antibodies. Soluble wild-type Pr78 (lane 1); pelletable wild-type Pr78 (lane 2); untransfected (lane 3); soluble Δ KKPKR Pr78 (lane 4); pelletable Δ KKPKR Pr78 (lane 5). (B) Virus release kinetics. Transfected COS-1 cells pulsed labeled with [³⁵S] methionine-cysteine for 30 minutes and chased for 0, 1, 2, 4, and 8 hours. Untransfected (lane 1); pSARM-4 (lanes 2–6); Δ KKRKR (lanes 7–11). Medium was collected and cells were lysed at the appropriate times with 1 × Buffer A. Cellular lysates and medium were adjusted to 1 × lysis buffer B. Viral proteins were immunoprecipitated from all samples using rabbit anti-Pr78 antibodies and separated by SDS-PAGE (12% acrylamide) and detected by phosphor-imaging.

(Fig. 2B). The 68 kDa protein is a N-terminal truncated Gag protein that is expressed by initiation of translation at an internal methionine codon of *gag* at position 100 [24]. As expected, the intensities of these wild-type Gag proteins in the cell lysates decreased during the chase with a concomitant appearance of p27 (CA). During or shortly after virus release, Pr78^{WT} is processed by the virus-encoded protease into p10 (MA), Np24, pp16/18, p12, p27 (CA), p14 (NC), and p4. Thus the appearance of p27 in the culture medium indicates that virus particles were

released and that Pr78^{WT} was being processed normally (Fig. 2B, lane 4). The other Gag cleavage products were not detected because they either do not contain methionines, or contain only a single methionine and thus were not detected.

In cells expressing the mutant virus, similar levels of cell-associated Gag precursors were observed in the pulse. Yasuda and Hunter [35] previously reported that deletion of the entire Np24 domain from Pr78 caused a rapid turn

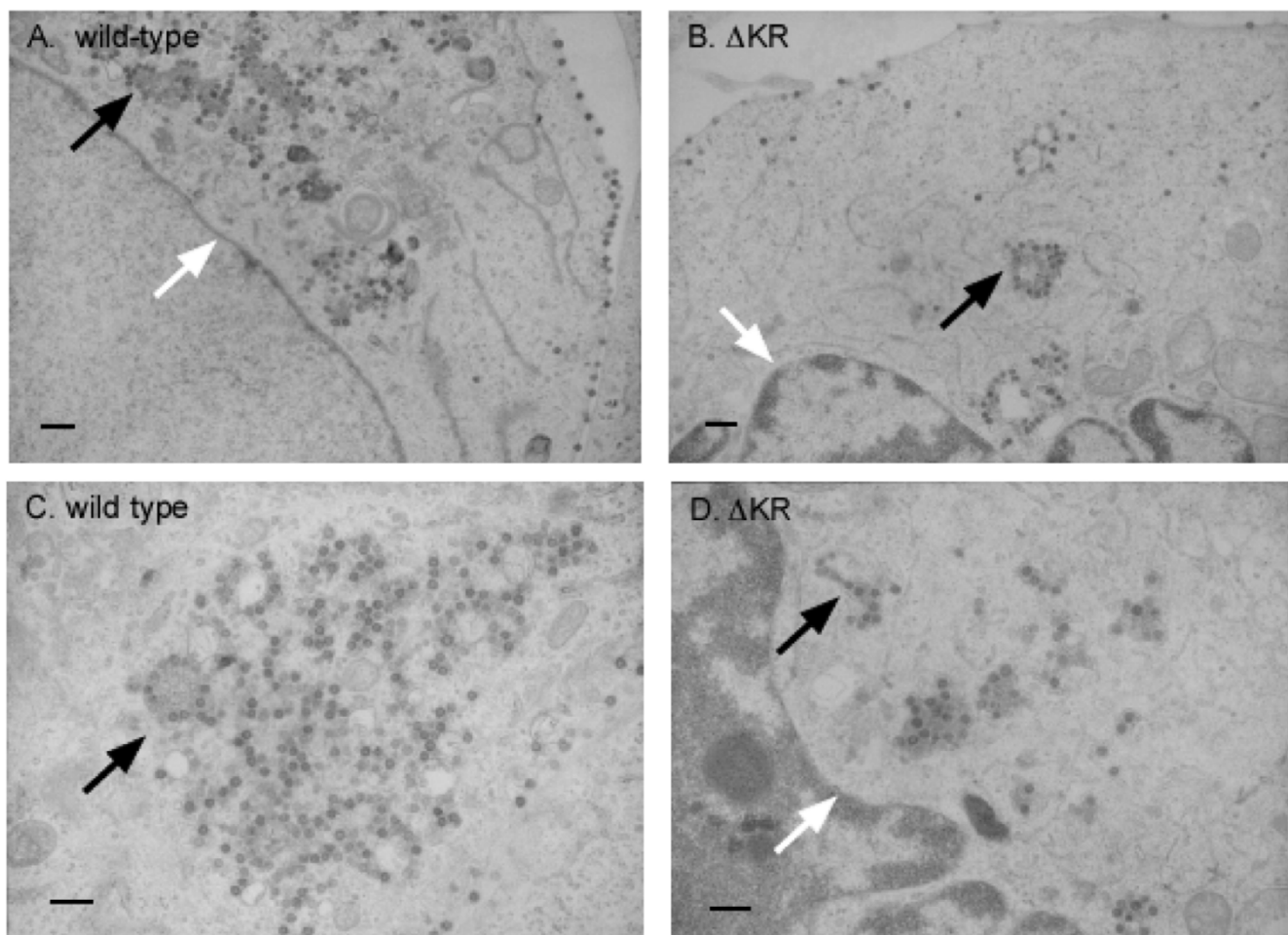


Figure 3

Intracellular procapsid morphology viewed by electron microscopy. COS-1 cells were transfected with pSARM-4 (A and C) or the Δ KKPKR mutant (B and D) proviral DNAs. Wild-type and mutant procapsids observed in close proximity to the nuclear membrane (white arrow) and throughout the cytoplasm near intracellular vesicles (black arrows). Bars approximately 500 nm.

over of Pr78 in cells and thus decreased the amount of particles released from cells. It was, therefore suggested that the Np24 domain is important for Pr78 stability. However, deletion of just the KKPKR motif did not alter the intracellular stability of Pr78^{ΔKR}. Instead, the intensity of Pr78^{ΔKR} decreased in the cell lysates in a manner similar to Pr78^{WT}. This was accompanied by a slightly reduced rate of release of virus particles; Pr78^{WT} can first be detected in the medium after 1 h (Fig. 3B), while Pr78^{ΔKR} was first detected in the medium after 2 h (Fig. 3B). Thus, Pr78^{ΔKR} efficiently assembled into procapsids and released processed Gag (p27) in the culture medium with kinetics similar to Pr78^{WT}.

ΔKKPKR intracellular procapsids are indistinguishable from WT procapsids

The metabolic labeling and cell fractionation experiments provided biochemical evidence that the deletion of the

KKPKR motif did not affect the ability of Gag to assemble into procapsids and be released from cells. Examining cells expressing Pr78^{WT} and Pr78^{ΔKR} by thin-section EM provided further evidence of normal assembly. Both produce intracellular, spherical procapsids (70–90 nm dia.) that have the characteristic, ring-shaped core typical of immature particles (Fig. 3). In addition, wild-type procapsids were found near the nuclear membrane, which others have shown to be the site of intracellular assembly [19] and near intracellular membranes (Fig. 3A and 3C). Interestingly, Sfakianos and Hunter have previously shown that Pr78^{WT} co-localizes with Rab11+ recycling endosomes [33]. Whether the vesicles shown here are recycling endosomes is not yet known. Of note, we observed fewer assembling, or fully assembled procapsids near the nuclear membrane compared to wild-type (Fig. 3B and 3D) suggesting that at least part of the replication defect

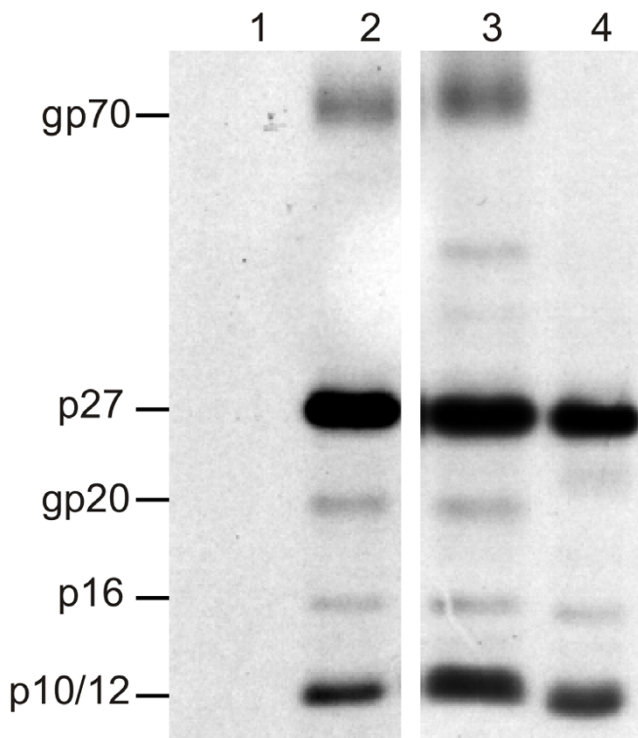


Figure 4
Glycoprotein incorporation and Gag processing. COS-1 cells were transfected with nothing (lane 1), pSAMR4 (lane 2), Δ KKPKR (lane 3), or pMT Δ E (lane 4) and then labeled overnight with [3 H] leucine. Culture medium was filtered and viral particles were pelleted through a 20% sucrose cushion. The viral proteins present in the pellet were immunoprecipitated using goat anti-M-PMV antibodies. Positions of various viral proteins are indicated.

may be due to defect in intracellular targeting of newly synthesized Pr78^{AKR} proteins.

Δ KKPKR Gag maturation and Envelope packaging

The cell fractionation and pulse-chase experiments combined with the EM analyses showed that the Δ KKPKR deletion mutant was released from cells as virus-like particles. Furthermore, the presence of reverse transcriptase activity and the CA (p27) protein in the culture medium of Δ KKPKR transfected cells suggest that the deletion did not affect PR-mediated processing of Gag, Gag-Pro or Gag-Pro-Pol (Fig. 1 and 2). The other Gag cleavage products (MA, p24/pp18, p12, NC, and p4) were not detected in the [35 S] methionine labeling experiments because they do not contain a sufficient number of methionine residues for detection. Furthermore, these experiments could not determine if the viral glycoproteins, SU and TM (gp70 and gp20, respectively) were incorporated into the released particles because an anti-pr78-specific antibody used for the immunoprecipitations. It was therefore pos-

sible that the KR box deletion mutation either inhibited Env glycoprotein incorporation or precise Gag processing, thus blocking infectivity. To address this possibility, we looked for the presence of the Env glycoproteins and the other Gag cleavage products in released virions. To this end, COS-1 cells were transfected with pSAMR4, p Δ KKPKR or an Env-deletion mutant, pMT. Δ E and labeled overnight with [3 H] leucine. Viral particles were pelleted through a 25% sucrose cushion, lysed, and immunoprecipitated using an anti-M-PMV antisera that recognizes the Gag cleavage products as well as gp70 and gp20. Figure 4 shows both wild-type and Δ KKPKR mutant particles contain the Env glycoproteins (gp70 and gp20) and the mature Gag cleavage proteins (MA [p10], pp16, p12, and CA [p27]). The NC (p14) and p4 cleavage products were not detected with the antiserum used. As expected particles released from pMT. Δ E transfected cells did not contain the gp70 or gp20 glycoproteins. These results demonstrate that the block to Δ KKPKR replication is not due to abnormal Pr78^{AKR} processing or an inability of the mutant particles to incorporate the viral glycoproteins.

Genome packaging

Having demonstrated that the deletion of the KR box did not affect the packaging or processing of the gag-, pol-, and env-encoded viral proteins, semi-quantitative RT-PCR assays were utilized to address whether the deletion of the KR box altered packaging of genomic RNA into virions. Equivalent amounts of virus, normalized by p27 content from wild-type and mutant virions were pelleted through a 20% sucrose cushion and resuspended in PBS and the viral RNAs were extracted. Two-fold serial dilutions of viral RNAs were used for RT-PCR reactions using primers to amplify CA-coding sequences. The relative amounts of viral RNA that were packaged were estimated by determining the end-point dilution within which viral cDNAs could be detected by ethidium bromide staining. As shown in this representative experiment, the deletion of the KKKPKR motif resulted in a 6–8 fold decrease in genome packaging, relative to wild-type (Fig. 5). Similar results were found using northern blot and dot-blot analyses of vRNAs using a riboprobe specific for M-PMV LTR sequences (data not shown). We concluded from these vRNA packaging assays that the deletion of the KR box significantly reduced the efficiency of vRNA packaging.

Subcellular localization

Electron microscopic examination indicated that fewer numbers of Δ KKPKR procapsids were present near the nuclear membrane and suggested that the deletion of the KR box influences intracellular targeting of Pr78^{AKR} to this perinuclear site of assembly. This observation combined with the finding that Pr78^{AKR} packages significantly less vRNA into particles lead us to hypothesize that correct

intracellular targeting and vRNA packaging are linked. This hypothesis is supported by previous studies with Rous sarcoma virus (RSV) and bovine leukemia virus (BLV) which demonstrated that basic residues in the regions of Gag proteins distant from the RNA binding motif within NC influences both Gag targeting and viral RNA packaging [29,30,32,36].

Studies with RSV have shown that its Gag proteins cycles through the nuclear compartment using a nonclassical nuclear targeting sequence within the MA domain and is exported out of the nucleus via the CRM-1 export pathway. Scheifele *et al.* have shown that treating the RSV Gag-expressing cells with the CRM-1 inhibitor leptomycin B (LMB) results in a dramatic accumulation of RSV Gag proteins within the nucleus. In addition, it has been shown that RSV MA mutants that are not targeted to the nuclear compartment are insensitive to LMB treatment and are released from cells as virus-like particles, yet are not infectious due to a defect in vRNA packaging. These results suggests that nuclear localization of RSV Gag and genome packaging are linked [29,36]. Likewise, Wang *et al.* have shown that basic residues within the MA domain of BLV are involved in vRNA packaging [30]. However, BLV Gag was not detected in the nucleus of cells treated with LMB. These results suggest that BLV Gag either does not enter the nucleus or that Gag does enter the nucleus but is exported by a CRM1-independent pathway.

To further explore the intracellular trafficking of Pr78^{WT} and Pr78^{AKR}, the steady-state intracellular locations of both were analyzed by confocal microscopy. Figure 6, which are representative z-sections of transfected COS-1 cells, shows that the highest concentration of both Pr78^{WT} and Pr78^{AKR} were routinely found throughout the cytoplasm. Interestingly, small amounts of Pr78^{WT} were also observed associated with the nuclear compartment. In contrast, nuclear staining of Pr78^{AKR} was only occasionally observed (Figure 6A and 6B, respectively). To examine if M-PMV Gag transiently traffics through the nuclear compartment in a CRM-1-dependent manner similar to RSV, we asked whether M-PMV Gag could be trapped within the nucleus by inhibiting the CRM-1 nuclear export pathway with LMB. As has been previously described [36], RSV Gag-GFP proteins were readily concentrated within the nucleus upon treatment with LMB (Fig. 6E-F). In contrast, LMB did not concentrate either Pr78^{WT} or Pr78^{AKR} in the nucleus (Fig. 6A-D).

Although these immunofluorescence experiments demonstrated that M-PMV does not utilize a CRM-1-dependent nuclear export pathway, it is possible that M-PMV Gag either enters the nucleus, and is exported in a CRM-1-independent manner, or Gag does not enter the nucleus but instead localizes to the nuclear pores. To distinguish

between these two possibilities, we utilized confocal microscopy, an affinity-purified rabbit polyclonal anti-Pr78 antibody and a monoclonal antibody (MAB414) that recognizes conserved FG repeats of nuclear pore proteins [37] to examine whether Gag co-localizes with nuclear pores. Following confocal imaging of HeLa cells expressing either Pr78^{WT} or Pr78^{AKR}, 0.3 μ m optical z-sections were stacked and orthogonal views through the nuclei were analyzed. In these cells, the nuclear pores were easily identifiable as punctate staining areas on the nuclear membrane. Moreover, Pr78^{WT} was not randomly dispersed within the nuclear compartment or around the nuclear membrane but rather concentrated in distinct foci in close proximity to nuclear pores (Fig. 6G-J). In contrast Pr78^{AKR} did not readily associate with nuclear pores (Fig. 6K-N), but instead localized mainly in the cytoplasm and as discrete foci adjacent to, but not associated with nuclear pores. However, we occasionally, but very infrequently, observed Pr78^{AKR} associating with nuclear pores.

Discussion

The results of the studies described here show that a KKPKR sequence located near the carboxy-terminal end of the M-PMV Np24 domain of Gag plays a critical role during virus replication. Initial experiments showed that deletion of this motif did not inhibit the release of virions as demonstrated by the release of virion-associated RT activity into the culture medium of transfected cells. However, this mutant was unable to replicate, similar to what was observed when the entire Np24 domain was deleted [35]. While it is possible that the replication defect was due to the deletion inducing deleterious conformational changes in Pr78, several observations argue that this is unlikely. First, the mutant was capable of assembling into spherical procapsids with morphologies indistinguishable from wild-type. Second, these mutant procapsids, like wild-type, associated with intracellular membranes as has been described as a normal transport pathway for several retroviruses [33,38,39]. Finally, they packaged normal levels of the viral glycoproteins, gp70 and gp20, into the released virions. These results show that the deletion did not significantly alter the confirmation of Pr78 and they demonstrate that the basic residues in Np24 are not involved in targeting Gag to cellular membranes or in packaging the viral glycoproteins.

Further analyses showed that the deletion resulted in two assembly-related defects, vRNA packaging and intracellular targeting. Semi-quantitative PCR, northern blot and dot blot analyses routinely demonstrated that the mutant packaged 6–8 fold less vRNA than did wild-type. In vitro assembly assays have shown that assembly of RSV and HIV-spherical capsids requires nucleic acids [26,40]. In the absence of nucleic acids, Gag proteins assemble into sheets and tubes. Although we have not yet determined

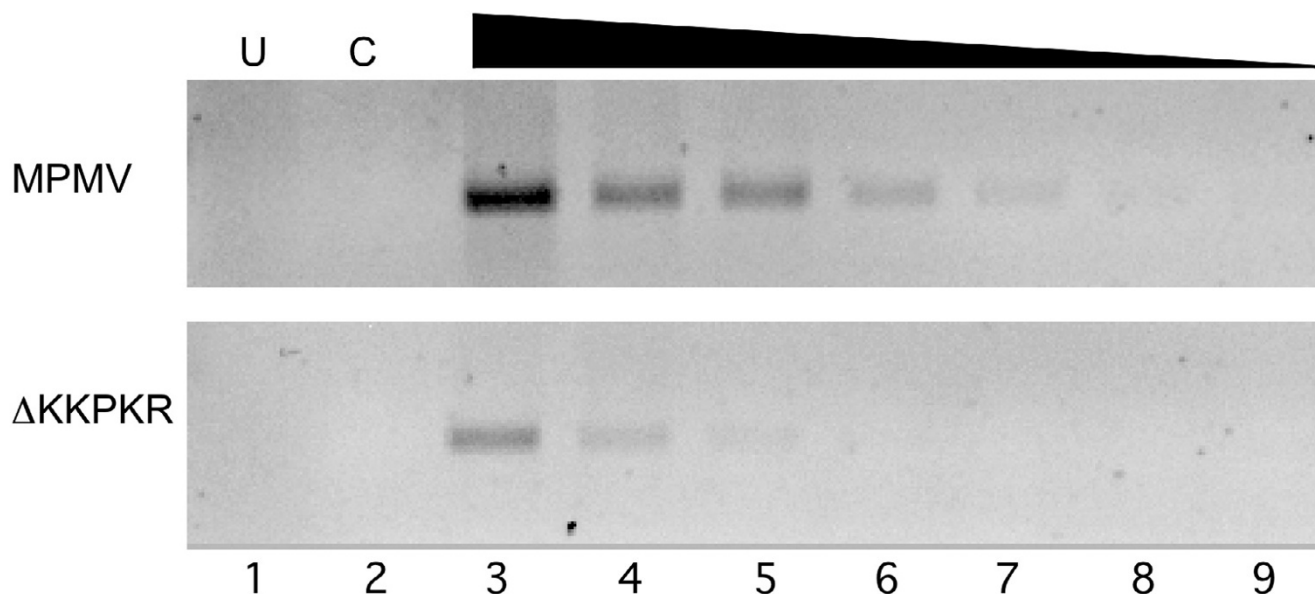


Figure 5

RT-PCR analysis of genome packaging in wild-type M-PMV and Δ KKPKR virions. Purified RNA from equivalent amounts of virus was diluted 1:1,000 (lane 3) followed by 2-fold serial dilutions to 1:96,000 (lane 9). First-strand cDNA synthesis was carried out using M-MLV RT and followed by PCR using oligos that amplify M-PMV CA sequences. Relative viral RNA packaging efficiencies were estimated by determining the end-point dilution in which viral PCR products could be detected by Ethidium bromide staining. U, untransfected (lane 1); C, RNA control – no reverse transcriptase added to RT-PCR reaction (lane 2).

whether the spherical, cytoplasmic procapsids assembled from the Δ KKPKR mutant contained RNA (presumably mainly cellular RNAs), we would assume that the RNA content of those intracellular capsids is similar to that found in the released capsids.

Several explanations could account for the failure of this mutant to specifically package vRNAs. First, the Np24 domain may be directly involved in RNA packaging. While there is no evidence that the Np24 domain directly binds RNA or interacts with the NC protein in the virion, the presence of these basic residues may help facilitate NC-mediated vRNA packaging in a manner analogous to that hypothesized for the MA domain of bovine leukemia virus [30].

Another explanation for the defect in genome packaging, although unlikely, is that the deletion disrupted the viral RNA dimerization and/or packaging signals. While it has been proposed that dimer formation is required for RNA packaging, the relationship between dimer formation and packaging is still unclear. Nonetheless, for RSV, MLV, and HIV, the sequence elements involved in dimerization are included in the packaging signals located near the 5'-end of their respective *gag* genes. In addition, many in vivo studies have shown that mutations that disrupt RNA dimer formation also interfere with RNA packaging. For

M-PMV, the RNA dimer initiation sequence (DIS) has not been precisely mapped. However, based on the mapping of the DIS within RNA packaging signals in other retroviruses (reviewed in [41]), the M-PMV DIS is also likely to be an integral part of the RNA packaging signal (Ψ). Because the Δ KKPKR deletion is more than 400 nucleotides downstream of Ψ packaging signal [42], it is unlikely that the deletion mutation affected the cis-acting elements required for vRNA dimerization or packaging. We are currently determining if the few viral RNAs detected in the Δ KKPKR particles exist as dimers and whether the mutant vRNA can be packaged in trans with wild-type Gag.

Previous studies on M-PMV suggested that the Np24 domain is required for Pr78 stability. Yasuda and Hunter [35] showed using pulse-chase experiments that deletion of the entire Np24 domain resulted in a Gag protein that when expressed in transfected cells, was more unstable than wild-type. The data presented here suggests that Np24, and perhaps more specifically the KKPKR motif, also plays an important role in intracellular targeting of Pr78. Based on the observations that relatively few Δ KKPKR mutant capsids were found in the perinuclear region of the cytoplasm, which has been shown to be the site of assembly, as well as the findings that wild-type Gag, but not the mutant, localized with the nuclear pores, we

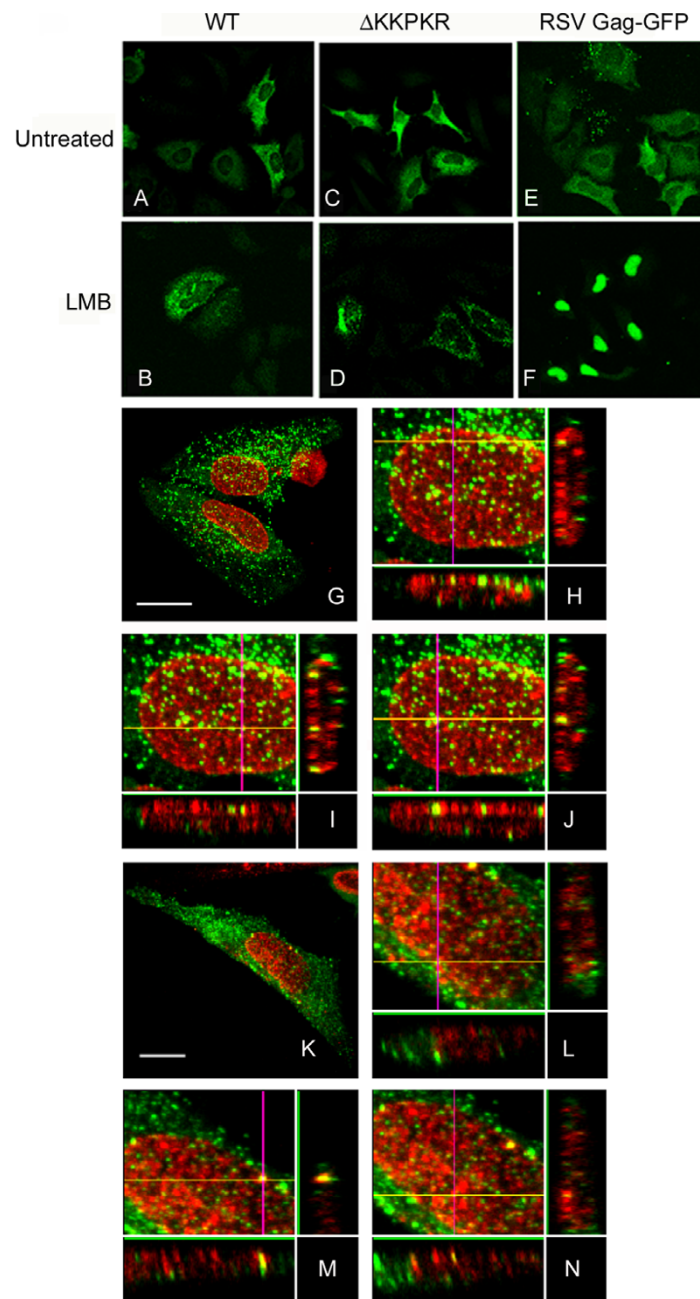


Figure 6

Subcellular localization of wild-type M-PMV Gag, Δ KKPKR Gag, and RSV Gag-GFP under steady state growth conditions or after treatment with LMB. HeLa cells were transfected with either pSARM-4, Δ KKPKR, or RSV Gag-GFP and left untreated or treated with LMB. The cells were fixed in methanol and the subcellular localizations of Gag were viewed by confocal microscopy using rabbit anti-Pr78 antibodies and Cy2 conjugated secondary antibodies. RSV Gag-GFP was directly visualized by fluorescence of the Gag-GFP fusion protein. Drug treatments: Wild-type M-PMV (untreated, panel 6A), wild-type MPMV (LMB treated, panel 6B) Δ KKPKR (untreated, panel 6C, Δ KKPKR (LMB treated, panel 6D), RSV Gag-GFP (untreated, panel 6E), and RSV Gag-GFP (LMB treated, panel 6F). Colocalization of wild-type M-PMV Gag, Δ KKPKR, and nuclear pores. Transfected HeLa cells were fixed with 4% paraformaldehyde, and permeabilized with 0.2% TX-100. Wild-type Gag (panels G-J), Δ KKPKR Gag (panels K-N), and nuclear pore localization were visualized by confocal microscopy using affinity purified anti-Pr78 and MAb414 antibodies, respectively, and counter stained with Cy2 anti-rabbit and Cy5 anti-mouse antibodies. 0.3 μ m Z-sections were stacked and orthogonal views through the cell were generated using Flowview imaging analysis software.

hypothesize that the KKPGR motif plays a role in targeting Gag to the site of genome packaging, which may be either be at the nuclear pores, within the nucleus, as suggested by Scheifele et al. [36], or in the cytoplasm juxtaposed to the outer nuclear membrane.

Interestingly, the KKPGR motif resembles a classical nuclear localization signal [43] and we have previously found that over expression of the nuclear pore associated, Ubc9 protein lead to a dramatic redistribution of Pr78 to the nuclear compartment [44]. Experiments are in progress to determine whether the KKPGR motif can function as a nuclear targeting signal when fused to a heterologous protein. If the KKPGR does function as an NLS to cycle Pr78 through the nucleus during the virus life cycle, as has been suggested for RSV, its export to the cytoplasm does not utilize the CRM-1 pathway (Fig. 6B). An alternative hypothesis, which is consistent with the data presented here, is that the KKPGR sequence targets Pr78 to the nuclear pore, where it first recognizes the vRNA during Tap-mediated RNA export [45]. This Gag-vRNA complex would then serve as the nucleation event for spherical capsid assembly just outside of the nuclear pore where the betaretroviruses are known to assemble.

A motif within the M-PMV MA domain called the cytoplasmic targeting/retention signal (CTRS), which is located approximately 100 residues upstream of the KKPGR motif, has also been implicated in directing Pr78 to the intracellular site of assembly. Mutant Pr78 proteins (R55W) that contain an arginine to tryptophan substitution at position 55 in MA do not accumulate at the usual cytoplasmic sites of assembly. Instead R55W-Pr78 proteins are targeted to the plasma membrane where they assemble concomitantly with budding, as with the C-type retroviruses [20]. This arginine is contained within an 18 amino acid sequence (residues 43–60) that is conserved between M-PMV and MMTV. When these 18 residues were inserted in the MA domain of C-type MLV Gag, MLV capsid assembly occurred in the cytoplasm [46]. Whether or not these altered MLV capsids assembled in the perinuclear/pericentriolar region of the cytoplasm was not shown. It has, therefore, been suggested that these residues either target Pr78 molecules to the cytoplasmic assembly site or they retain Pr78 at this site until the pro-capsids are fully assembled.

We hypothesize that the KKPGR motif identified in this study, which may be included in a larger motif that has yet to fully defined, functions either prior to or separate from the CTRS function. We speculate that the KKPGR motif is involved in targeting Pr78 to the nuclear pore to facilitate RNA packaging. Because only two copies of vRNA are packaged into virions, only a few Gag proteins need to be targeted there, which is consistent with the findings

present here that the majority of Gag proteins don't associate with nuclear pores. Once Gag has associated with the vRNA, the Gag-vRNA complex would then be transported to the assembly site perhaps via the CTRS signal to initiate spherical capsid assembly.

Materials and methods

DNAs

Plasmid pSARM4 is an infectious molecular clone of wild-type M-PMV. Plasmid pMT.ΔE is an env deletion mutant of pSARM4 [47]. Deletion of the KKPGR motif was accomplished using the Altered Sites II Mutagenesis System (Promega) as per manufacture's protocol. Briefly, the 1,307 bp, SphI-PvuII fragment (nt 171-1478) of pSARM4 was subcloned into the SmaI-SphI sites of pALTER. Mutagenesis was carried out using the mutagenic oligonucleotide (5'-GTTTGTGCTCTTAACAGAACTGGGAAAGTACTTGATAAACCTTTATCTTGTAGAGAGG), to precisely delete amino acids 153 through 157 (KKPGR) in M-PMV Gag. The mutation was subcloned back into pSARM4 using SacI and PaeI sites. After mutagenesis, plasmid DNAs were sequenced to ensure that unwanted mutations were not inadvertently created. Plasmid pETM100A is a prokaryote expression vector used to express a (His)₆-tagged M-PMV Gag protein in *E. coli* (32). Plasmid pRS.V8-EGFP was used to express a RSV Gag-EGFP fusion protein in mammalian cells (John Wills, Pennsylvania State University College of Medicine) [29].

Cell lines and transfection

COS-1 and HOS cells were grown at 37°C with 5% CO₂ in Dulbecco's modified Eagle's medium supplemented with 10% fetal bovine serum. HeLa cells were grown in RPMI 1640 medium supplemented with 10% fetal bovine serum and 5% tryptose phosphate broth. DNA transfections were carried out using Fugene 6 (Roche Diagnostics, Indianapolis, IN) following the manufacture's protocol.

Antibodies

Goat anti-M-PMV antibodies were obtained from Eric Hunter (Emory University). Mouse monoclonal antibodies that recognize the conserved FG repeats found in nuclear pore complex proteins (Mab414) were purchased from Covance Research Products (Berkely, CA). HRP-conjugated goat anti-rabbit IgG and HRP-conjugated goat anti-mouse IgG were purchased from Amersham Pharmacia Biotech (Little Chalfont Buckinghamshire, England). Cy2 conjugated donkey anti-rabbit and Cy5 conjugated donkey anti-mouse were purchased from Jackson ImmunoResearch Laboratories (West Grove, PA). Rabbit polyclonal anti-Pr78 (47) was affinity purified as follows. M-PMV Gag proteins containing a carboxy-terminal (His)₆ tag were expressed in *E. coli* BL21 (DE3) cells from the expression plasmid pET.M100A for 4 hours in the presence of 0.1 mM IPTG. Cells were harvested by centrifuga-

tion at $4,000 \times g$ and lysed at room temperature in a denaturing lysis buffer containing 8 M urea in TNI pH 8.0 buffer (50 mM Tris Cl pH 8.0, 150 mM NaCl and 0.5 mM imidazole). Cellular debris was removed by centrifugation at $10,000 \times g$ and the supernatant was passed over an Ni-NTA agarose nickel column (Qiagen Sciences, Maryland, USA) and extensively washed in 8 M urea in TNI pH 6.5 buffer. The denatured Gag proteins on the column were refolded using a slow (8 h), linear reduction of urea from 8 M to 0 M (in TNI buffer, pH 8.0) in a manner similar to that described by Klikova et al. [48] who have shown that M-PMV Gag proteins denatured in 8 M urea could be refolded into a confirmations that are competent to assemble into immature capsids in vitro by slowly removing the urea over an 8 hour period at 4°C . After refolding, Gag proteins were eluted from the Ni^{2+} column using TNI pH 8.0 buffer containing 0.1 mM EDTA. After exchanging the TNI-EDTA buffer to pH7.2 Coupling Buffer (Pierce) by dialysis, the Gag proteins were covalently coupled to activated agarose beads using the Amino-Link Plus kit (Pierce, Rockford, IL) per manufacturer's protocol. Anti-Pr78 antibodies in the immunized rabbit serum were affinity purified using the immobilized Gag column by standard protocols [49].

Intracellular Gag Fractionation and Immunoblots

Intracellular Gag fractionation experiments were used to assay intracellular procapsid assembly. 60 mm diameter culture dishes containing either untransfected or transfected COS-1 cells were lysed in 1 ml TX-100 lysis buffer (0.25 M sucrose, 1.0 mM EDTA, 10 mM Tris-HCl [pH7.5], 0.14 M NaCl, 0.5% Triton X-100, 0.25% DOC) containing PMSF, leupeptin, pepstatin, and aprotinin for 30 minutes on ice. Cell debris and nuclei were removed by centrifugation for 2 min in a microfuge. The pellets were discarded and the capsid-containing supernatants were fractionated by centrifugation through a 20% sucrose cushion at $350,000 \times g$ in a MLA-130 rotor for 30 minutes at 4°C . The pelleted procapsids were lysed in $2\times$ protein loading buffer (10% glycerol, 2.3% SDS, 63 mM Tris-HCL [pH6.8], 5% β -mercaptoethanol, and 0.01% bromophenol blue). Lysates were boiled for 3 minutes, separated on a 12% SDS PAGE gel, transferred to nitrocellulose. Viral proteins were analyzed by western blot using polyclonal anti-Pr78, HRP-conjugated goat anti-rabbit IgG, and Western Lightning Chemiluminescence Reagent Plus (Perkin-Elmer) as per manufacturers suggestions.

Radiolabeling and immunoprecipitation

For this, subconfluent 60 mm diameter tissue culture dishes containing either untransfected and transfected COS-1 cells were washed twice in DMEM without methionine or cysteine (DMEM Met⁻ Cys⁻) and starved in 4 ml of DMEM Met⁻ Cys⁻ for 15 min at 37°C in 5% CO_2 . The medium was replaced with 800 μl of DMEM Met⁻ Cys⁻

containing 250 μCi [^{35}S] methionine-cysteine (>1000 Ci/mmol; NEN, Boston, MA). The cells were incubated for 30 minutes at 37°C , 5% CO_2 . Cells were either immediately lysed (pulse) or washed with complete medium and then incubated for 1, 2, 3, 4, or 8 hours in complete medium (chase) prior to lysis.

Virion and cell lysis were lysed as follows. Culture medium from each plate was collected and clarified by centrifugation in a microfuge for 2 min at 13,000 rpms. Virions were lysed by adjusting the culture medium to $1\times$ lysis buffer B by adding 1/5 volume of $5\times$ lysis buffer B (0.5% sodium dodecyl sulfate [SDS], 5% Triton X-100, 5% deoxycholate [DOC], 0.75 M NaCl, 0.25 M Tris-HCl [pH 6.8]). Cell monolayers were lysed in 1 ml lysis buffer A (Triton X-100, 1% DOC, 0.15 M NaCl, 0.05 M Tris-HCl [pH 6.8]) containing PMSF, leupeptin, pepstatin, and aprotinin for 30 minutes on ice. Cell debris and nuclei were removed by centrifugation for 2 min in a microfuge. The pellets were discarded and the capsid-containing supernatants were adjusted to $1\times$ lysis buffer B by adding 0.1% SDS. All lysates were precleared by incubation with inactivated, formalin-fixed *Staphylococcus aureus* cells. Viral proteins were immunoprecipitated from all samples using Rbt anti-Pr78 antibodies and *Staphylococcus aureus* cells as previously described [50]. Immunoprecipitates were resuspended in protein loading buffer (10% glycerol, 2.3% SDS, 63 mM Tris-HCL [pH6.8], 5% β -mercaptoethanol, and 0.01% bromophenol blue), boiled for 3 minutes, separated by SDS PAGE gel (12% acrylamide), and visualized by fluorography or analyzed by phosphor-imaging using The Discovery Series Quantity One (Bio-Rad, Hercules, CA).

Steady state radiolabeling with [^3H] leucine was done to assess glycoprotein incorporation and Pr78 processing. Transfected COS-1 cells were starved in leucine-free DMEM for 90 minutes, and labeled overnight with [^3H] leucine (500 $\mu\text{Ci}/\text{ml}$, 173.0 Ci/mmol, NEN, Boston, MA). The culture medium was filtered with a 0.45 μm syringe filter, and [^3H] leucine-labeled viruses were pelleted through a 25% sucrose cushion by centrifugation at $350,000 \times g$ in a TLA 100.3 rotor for 30 minutes at 4°C . After the viral pellet was solublized in $1\times$ lysis buffer B, the viral proteins were using goat anti-M-PMV antibodies and analyzed as described above.

Virus replication assay

The infectivity of wild-type and mutant viruses were determined by measuring the increase of reverse transcriptase (RT) activity in the culture supernatants of inoculated HOS cell cultures at various times postinfection. Culture medium was harvested from COS-1 cells that had been transfected 48 h previously with either wild-type or mutant DNA. Loose cells and cellular debris were pelleted

by centrifugation for 2 min in a microfuge and the level of RT activity in the clarified culture medium was measured. Hos cells were infected with equivalent amount of RT-containing in the presence of 4.0 µg/ml of polybrene. Culture fluids were harvested at various days postinfection and assayed for RT activity. RT assays were carried out by pelleting virus from medium by centrifugation at 350,000 × g in a TLA 100.3 rotor for 30 minutes at 4°C. The viral pellet was lysed in 12 µl of Virion Lysis Buffer (50 mM Tris-HCl [pH 7.8], 100 mM KCl, 0.05% TX-100, 2 mM dithiothreitol [dT]) on ice for 15 min. 7.5 µl of virion lysates were added to 30 µl of RT Reaction Buffer (50 mM Tris [pH 8.0], 100 mM KCl, 2 mM dTT, 7.5 mM MgCl₂) with 8 µCi [³²P] α-TTP (>1000 mCi/mmol, NEN) and 1.25 µg poly (A) – oligo (dT)₁₅ (Roche). The RT reaction was incubated at 37°C for 2 hours. 10 µl of the RT reaction was spotted onto a piece of Whatman DE81 paper and allowed to dry. The filter paper was washed twice for 15 min in 2× SSC buffer (0.3 M NaCl, 0.03 M Na Citrate), twice briefly in 95% EtOH, and once in distilled water. The filters were dried and [³²P] incorporation was measured by scintillation counting.

RNA extraction and RT-PCR

Medium from transfected COS-1 cells were clarified and virus was pelleted through a 20% sucrose cushion at 207,570 × g in an SW41 rotor for 2 hours at 4°C and resuspended in 30 µl of PBS. The amount of viral particles was normalized by quantitation of p27 detected by immunoblotting. RNA was extracted from equivalent amounts of virus using QIAamp Viral RNA Mini Kit (Qiagen Sciences, Maryland, USA) as per manufacturers suggestions. Purified RNA was treated with 1 U of Rnase-free Dnase I (New England Biolabs, Inc., Beverly, MA) for 30 min at 37°C, followed by inactivation at 70°C for 30 min. Purified RNA from equivalent amounts of virus was diluted 1:1,000 followed by 2-fold serial dilutions. 5 µl of diluted RNA was used for first-strand cDNA synthesis as per manufacturers suggestions for M-MLV RT (Invitrogen) using 500 ng of oligo (dT)¹²⁻¹⁸ (Ambion, Austin, TX). First-strand cDNA (5 µl) were amplified by PCR using the following oligos 5'-CCGCTCGAGCGGGCCGCCATGCCGGTGGCTGAAACCGTTG and 5'-GCTCTAGAGCGCGGCCATGGCCAGG to amplify M-PMV CA sequences. The relative amount of viral RNA packaged into viral particles was estimated by end-point dilution.

Electron microscopy

Transmission Electron Microscopy (TEM) was utilized to view assembled intracellular procapsids. Transfected COS-1 cells were fixed for 1 hour with two changes of 3% glutaraldehyde in 0.1 M phosphate buffer (5 mM NaH₂PO₄ and 5 mM phosphoric acid). The cells were rinsed for 30 min with 0.1 M phosphate buffer, followed by osmication (2% OsO₄ in phosphate buffer) for 1 hour.

The cultures were washed and dehydrated with a graded series of ethanol (25% 50%, 75%, 95% 100), followed by a graded series of ethanol/Epon 812 (Shell) mixtures (3:1, 1:1, 1:3). The cells were infiltrated with pure Epon 812 and polymerized at 60°C for 48 hours. Thin sections were made with a LBK Ultratome III, mounted on copper grids, and stained with 2% uranyl acetate and lead citrate. Thin sections were examined and photographed with a HitachiH-7500 transmission electron microscope operated at 60 Kv.

Subcellular localization

The intracellular localizations of Pr78^{Gag} and Pr78^{ΔKR} proteins were determined by confocal microscopy. Transfected HeLa cells were grown on sterile coverslips in 35 mm culture dishes. At 48 hours post transfection, cells were washed with PBS (137 mM NaCl₂, 2.7 mM KCl₂, and 8 mM Na₂HPO₄, 2 mM KH₂PO₄), fixed in either 4% paraformaldehyde in PBS for 20 minutes at room-temperature and then subsequently permeabilized with 0.2% TX-100 in PBS for 5 minutes at room-temperature, or fixed in 100% methanol at -20°C for 10 minutes. The coverslips were washed with PBS, and blocked with Blocking Buffer 1 (PBS, 0.2% tween 20, 0.4% fish skin gelatin [Sigma]) for 5 minutes, and then blocked with Blocking Buffer 2 (PBS, 0.2% tween 20, 2.5% goat serum [Sigma]) for 5 min. Affinity purified anti-Pr78 and MAb414 (in PBS, 0.2% tween 20, 2.5% goat serum) were incubated on the coverslips for 45 min at 37°C. The primary antibodies were removed and the coverslips were blocked as they had been previously. Cy2 and Cy5 conjugated secondary antibodies (in PBS, 0.2% tween 20, 2.5% goat serum) were incubated on the cover-slips for 30 min at 37°C and washed in PBS with 0.2% tween 20. The coverslips were mounted on slides using GEL-mount and analyzed by confocal microscopy (Olympus FV500 w/upright BX Olympus fluorescence microscope). 0.3 µm Z-sections were stacked and orthogonal views through the cell were generated using Flowview imaging analysis software (Olympus).

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References

1. Baillie GJ, van de Lagemaat LN, Baust C, Mager DL: Multiple groups of endogenous betaretroviruses in mice, rats, and other mammals. *J Virol* 2004, **78**(11):5784-5798.
2. Hook LM, Agafonova Y, Ross SR, Turner SJ, Golovkina TV: **Genetics of mouse mammary tumor virus-induced mammary tumors: linkage of tumor induction to the gag gene.** *J Virol* 2000, **74**(19):8876-8883.
3. Xu L, Sakalian M, Shen Z, Loss G, Neuberger J, Mason A: **Cloning the human betaretrovirus proviral genome from patients with primary biliary cirrhosis.** *Hepatology* 2004, **39**(1):151-156.

4. Power MD, Marx PA, Bryant ML, Gardner MB, Barr PJ, Luciw PA: **Nucleotide sequence of SRV-1, a type D simian acquired immune deficiency syndrome retrovirus.** *Science* 1986, **231(4745)**:1567-1572.
5. Sonigo P, Barker C, Hunter E, Wain-Hobson S: **Nucleotide sequence of Mason-Pfizer monkey virus: an immunosuppressive D-type retrovirus.** *Cell* 1986, **45(3)**:375-385.
6. Thayer RM, Power MD, Bryant ML, Gardner MB, Barr PJ, Luciw PA: **Sequence relationships of type D retroviruses which cause simian acquired immunodeficiency syndrome.** *Virology* 1987, **157(2)**:317-329.
7. Colcher D, Heberling RL, Kalter SS, Schlom J: **Squirrel monkey retrovirus: an endogenous virus of a new world primate.** *J Virol* 1977, **23(2)**:294-301.
8. Carlson J, Lyon M, Bishop J, Vaiman A, Cribiu E, Mornex JF, Brown S, Knudson D, DeMartini J, Leroux C: **Chromosomal distribution of endogenous Jaagsiekte sheep retrovirus proviral sequences in the sheep genome.** *J Virol* 2003, **77(17)**:9662-9668.
9. Cousens C, Minguillon E, Dalziel RG, Ortin A, Garcia M, Park J, Gonzalez L, Sharp JM, de las Heras M: **Complete sequence of enzootic nasal tumor virus, a retrovirus associated with transmissible intranasal tumors of sheep.** *J Virol* 1999, **73(5)**:3986-3993.
10. Kajikawa O, Dahlberg JE, Rosadio RH, De Martini JC: **Detection and quantitation of a type D retrovirus gag protein in ovine pulmonary carcinoma (sheep pulmonary adenomatosis) by means of a competition radioimmunoassay.** *Vet Microbiol* 1990, **25(1)**:17-28.
11. Baillie GJ, Wilkins RJ: **Endogenous type D retrovirus in a marsupial, the common brushtail possum (*Trichosurus vulpecula*).** *J Virol* 2001, **75(5)**:2499-2507.
12. Mager DL, Freeman JD: **Novel mouse type D endogenous proviruses and ETn elements share long terminal repeat and internal sequences.** *J Virol* 2000, **74(16)**:7221-7229.
13. Risteviski S, Purcell DF, Marshall J, Campagna D, Nouri S, Fenton SP, McPhee DA, Kannourakis G: **Novel endogenous type D retroviral particles expressed at high levels in a SCID mouse thymic lymphoma.** *J Virol* 1999, **73(6)**:4662-4669.
14. Chopra HC, Bogden AE, Zelljadt I, Jensen EM: **Virus particles in a transplantable rat mammary tumor of spontaneous origin.** *Eur J Cancer* 1970, **6(4)**:287-290.
15. Fine DL, Landon JC, Pienta RJ, Kubicek MT, Valerio MG, Loeb WF, Chopra HC: **Responses of infant rhesus monkeys to inoculation with Mason-Pfizer monkey virus materials.** *J Natl Cancer Inst* 1975, **54(3)**:651-658.
16. Marx PA, Maul DH, Osborn KG, Lerche NW, Moody P, Lowenstein LJ, Henrickson RV, Arthur LO, Gilden RV, Gravell M, et al: **Simian AIDS: isolation of a type D retrovirus and transmission of the disease.** *Science* 1984, **223(4640)**:1083-1086.
17. Stromberg K, Benveniste RE, Arthur LO, Rabin H, Giddens WEJ, Ochs HD, Morton WR, Tsai CC: **Characterization of exogenous type D retrovirus from a fibroma of a macaque with simian AIDS and fibromatosis.** *Science* 1984, **224(4646)**:289-282.
18. Freed EO: **HIV-1 gag proteins: diverse functions in the virus life cycle.** *Virology* 1998, **251(1)**:1-15.
19. Sfakianos JN, LaCasse RA, Hunter E: **The M-PMV cytoplasmic targeting-retention signal directs nascent Gag polypeptides to a pericentriolar region of the cell.** *Traffic* 2003, **4(10)**:660-670.
20. Rhee SS, Hunter E: **A single amino acid substitution within the matrix protein of a type D retrovirus converts its morphogenesis to that of a type C retrovirus.** *Cell* 1990, **63(1)**:77-86.
21. Rhee SS, Hunter E: **Amino acid substitutions within the matrix protein of type D retroviruses affect assembly, transport and membrane association of a capsid.** *Embo J* 1991, **10(3)**:535-546.
22. Rhee SS, Hunter E: **Structural role of the matrix protein of type D retroviruses in gag polyprotein stability and capsid assembly.** *J Virol* 1990, **64(9)**:4383-4389.
23. Strambio-de-Castillia C, Hunter E: **Mutational analysis of the major homology region of Mason-Pfizer monkey virus by use of saturation mutagenesis.** *J Virol* 1992, **66(12)**:7021-7032.
24. Sakalian M, Hunter E: **Separate assembly and transport domains within the Gag precursor of Mason-Pfizer monkey virus.** *J Virol* 1999, **73(10)**:8073-8082.
25. Zabransky A, Sakalian M, Pichova I: **Localization of self-interacting domains within betaretrovirus Gag polyproteins.** *Virology* 2005, **332(2)**:659-666.
26. Campbell S, Rein A: **In vitro assembly properties of human immunodeficiency virus type I Gag protein lacking the p6 domain.** *J Virol* 1999, **73(3)**:2270-2279.
27. Campbell S, Vogt VM: **Self-assembly in vitro of purified CA-NC proteins from Rous sarcoma virus and human immunodeficiency virus type I.** *J Virol* 1995, **69(10)**:6487-6497.
28. Dupont S, Sharova N, DeHoratius C, Virbasius CM, Zhu X, Bukrinskaya AG, Stevenson M, Green MR: **A novel nuclear export activity in HIV-1 matrix protein required for viral replication.** *Nature* 1999, **402(6762)**:681-685.
29. Callahan EM, Wills JW: **Link between genome packaging and rate of budding for Rous sarcoma virus.** *J Virol* 2003, **77(17)**:9388-9398.
30. Wang H, Norris KM, Mansky LM: **Involvement of the matrix and nucleocapsid domains of the bovine leukemia virus Gag polyprotein precursor in viral RNA packaging.** *J Virol* 2003, **77(17)**:9431-9438.
31. Parent LJ, Cairns TM, Albert JA, Wilson CB, Wills JW, Craven RC: **RNA dimerization defect in a Rous sarcoma virus matrix mutant.** *J Virol* 2000, **74(1)**:164-172.
32. Garbitt RA, Albert JA, Kessler MD, Parent LJ: **trans-acting inhibition of genomic RNA dimerization by Rous sarcoma virus matrix mutants.** *J Virol* 2001, **75(1)**:260-268.
33. Sfakianos JN, Hunter E: **M-PMV capsid transport is mediated by Env/Gag interactions at the pericentriolar recycling endosome.** *Traffic* 2003, **4(10)**:671-680.
34. Gottwein E, Bodem J, Muller B, Schmechel A, Zentgraf H, Krausslich HG: **The Mason-Pfizer monkey virus PPPY and PSAP motifs both contribute to virus release.** *J Virol* 2003, **77(17)**:9474-9485.
35. Yasuda J, Hunter E: **A proline-rich motif (PPPY) in the Gag polyprotein of Mason-Pfizer monkey virus plays a maturation-independent role in virion release.** *J Virol* 1998, **72(5)**:4095-4103.
36. Scheifele LZ, Garbitt RA, Rhoads JD, Parent LJ: **Nuclear entry and CRM1-dependent nuclear export of the Rous sarcoma virus Gag polyprotein.** *Proc Natl Acad Sci U S A* 2002, **99(6)**:3944-3949.
37. Davis LI, Blobel G: **Identification and characterization of a nuclear pore complex protein.** *Cell* 1986, **45(5)**:699-709.
38. Kramer B, Pelchen-Matthews A, Deneka M, Garcia E, Piguet V, Marsh M: **HIV interaction with endosomes in macrophages and dendritic cells.** *Blood Cells Mol Dis* 2005, **35(2)**:136-142.
39. Nydegger S, Foti M, Derdowski A, Spearman P, Thali M: **HIV-1 egress is gated through late endosomal membranes.** *Traffic* 2003, **4(12)**:902-910.
40. Campbell S, Vogt VM: **In vitro assembly of virus-like particles with Rous sarcoma virus Gag deletion mutants: identification of the p10 domain as a morphological determinant in the formation of spherical particles.** *J Virol* 1997, **71(6)**:4425-4435.
41. Paillart JC, Shehu-Xhilaga M, Marquet R, Mak J: **Dimerization of retroviral RNA genomes: an inseparable pair.** *Nat Rev Microbiol* 2004, **2(6)**:461-472.
42. Schmidt RD, Mustafa F, Lew KA, Browning MT, Rizvi TA: **Sequences within both the 5' untranslated region and the gag gene are important for efficient encapsidation of Mason-Pfizer monkey virus RNA.** *Virology* 2003, **309(1)**:166-178.
43. Silver PA: **How proteins enter the nucleus.** *Cell* 1991, **64(3)**:489-497.
44. Weldon RAJ, Sarkar P, Brown SM, Weldon SK: **Mason-Pfizer monkey virus Gag proteins interact with the human sumo conjugating enzyme, hUbc9.** *Virology* 2003, **314(1)**:62-73.
45. Braun IC, Rohrbach E, Schmitt C, Izaurralde E: **TAP binds to the constitutive transport element (CTE) through a novel RNA-binding motif that is sufficient to promote CTE-dependent RNA export from the nucleus.** *Embo J* 1999, **18(7)**:1953-1965.
46. Choi G, Park S, Choi B, Hong S, Lee J, Hunter E, Rhee SS: **Identification of a cytoplasmic targeting/retention signal in a retroviral Gag polyprotein.** *J Virol* 1999, **73(7)**:5431-5437.
47. Song C, Hunter E: **Variable sensitivity to substitutions in the N-terminal heptad repeat of Mason-Pfizer monkey virus transmembrane protein.** *J Virol* 2003, **77(14)**:7779-7785.
48. Kilikova M, Rhee SS, Hunter E, Ruml T: **Efficient in vivo and in vitro assembly of retroviral capsids from Gag precursor proteins expressed in bacteria.** *J Virol* 1995, **69(2)**:1093-1098.
49. Harlow ELD: **Antibodies; A laboratory manual.** Cold Spring Harbor, Cold Spring Harbor Laboratory Press; 1988:726.

50. Sakalian M, Parker SD, Weldon RAJ, Hunter E: **Synthesis and assembly of retrovirus Gag precursors into immature capsids in vitro.** *J Virol* 1996, **70(6)**:3706-3715.

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